

**Program**  
**Nano2Life Summer School**  
*Methods in Micro – Nanotechnology and Nanobiotechnology*  
**30 June – 10 July 2008**

<b>MONDAY , 30 JUNE – NCSR “DEMOKRITOS”</b>	
<b>09:00-09:30</b>	<b>Welcome</b>
<b>09:30-10:15</b>	<u>Lecture 1.1</u> : Cell biology principles - Part 1 (Dr Dimitris Mastellos)
<b>10:15-11:00</b>	<u>Lecture 1.2</u> : Structure of biological macromolecules (Prof. Elias Eliopoulos)
<b>11:00-11:15</b>	<b>Coffee Break</b>
<b>11:15-12:00</b>	<u>Lecture 1.2</u> : Structure of biological macromolecules (Prof. Elias Eliopoulos)
<b>12:00-13:00</b>	<u>Lecture 1.1</u> : Cell biology principles - Part 2 (Dr Dimitris Mastellos)
<b>13:00-14:00</b>	<b>Lunch break</b>
<b>14:00-15:30</b>	<u>Lecture 1.3</u> : Microelectronic Materials and Device Technology (Dr Spyros Gardelis)
<b>15:30-15:45</b>	<b>Coffee Break</b>
<b>15:45-17:15</b>	<u>Lecture 3.2</u> : Lab on chip devices: Principles, applications, opportunities (Invited: Dr Joel Rossier)
<b>17:15-18:00</b>	<u>Lecture 2.3.7</u> : Applied Bioinformatics in BioNanoTechnology (Dr Georgios Spyrou)
<b>TUESDAY , 1 JULY – NCSR “DEMOKRITOS”</b>	
<b>09:00-10:30</b>	<u>Lecture 2.3.1</u> : Gel-based protein analysis methods (Dr Antonia Vlahou)
<b>10:30-11:15</b>	<u>Lecture 2.3.2</u> : Non-gel based protein analysis methods (Dr Spiros D. Garbis)
<b>11:15-11:30</b>	<b>Coffee Break</b>
<b>11:30-12:15</b>	<u>Lecture 2.3.3</u> : Binding Assays and Immunosensors Part 1 - Binding assays (Dr Sotirios Kakabakos)
<b>12:15-13:00</b>	<u>Lecture 2.3.3</u> : Binding Assays and Immunosensors Part 2 - Immunosensors (Dr Christos Mastichiadis)
<b>13:00-13:45</b>	<u>Lecture 2.3.4</u> : DNA and Protein arrays: fabrication, detection and applications (Dr Panagiota Petrou)
<b>13:45-14:45</b>	<b>Lunch break</b>
<b>14:45-15:30</b>	<u>Lecture 2.3.6</u> : Introduction into Bioinformatics (Dr Sophia Kossida)
<b>15:30-17:00</b>	<u>Lecture 1.4</u> : Introduction to nanobiotechnology (Invited: Prof. Yossi Shacham-Diamand)
<b>17:00-17:15</b>	<b>Coffee Break</b>
<b>17:15-18:45</b>	<u>Lecture 2.2.7</u> : Imaging with Scanning Probes (AFM, STM, SNOM) (Invited: Dr Martin Bennink)

## WEDNESDAY , 2 JULY – ACADEMY OF ATHENS

<b>09:00-09:45</b>	<u>Lecture 2.2.8</u> : Fluorescence imaging and 3D image visualization using confocal microscopy (Dr Stamatis Pagakis)
<b>09:45-10:00</b>	<b>Coffee Break</b>
<b>10:00-12:00</b> <b>(shift 1)</b>	<u>Laboratory 2.2.4</u> : State of the art confocal microscopy of biological samples (Dr Stamatis Pagakis) <u>Laboratory 2.3.1</u> : Protein separation by two-dimensional electrophoresis (Dr Antonia Vlahou) <u>Laboratory 2.3.2</u> : Mass spectrometry (Dr Spiros D. Garbis) <u>Laboratory 2.3.3</u> : Fabrication of protein microarrays using nanoplotter (Dr George Tsangaris) <u>Laboratory 2.3.6</u> : Bioinformatics laboratory (Dr Sophia Kossida)
<b>12:00-12:15</b>	<b>Coffee Break</b>
<b>12:15-14:15</b> <b>(shift 2)</b>	<u>Laboratory 2.2.4</u> : State of the art confocal microscopy of biological samples (Dr Stamatis Pagakis) <u>Laboratory 2.3.1</u> : Protein separation by two-dimensional electrophoresis (Dr Antonia Vlahou) <u>Laboratory 2.3.2</u> : Mass spectrometry (Dr Spiros D. Garbis) <u>Laboratory 2.3.3</u> : Fabrication of protein microarrays using nanoplotter (Dr George Tsangaris) <u>Laboratory 2.3.6</u> : Bioinformatics laboratory (Dr Sophia Kossida)
<b>14:15-15:15</b>	<b>Lunch break</b>
<b>15:15-17:15</b> <b>(shift 3)</b>	<u>Laboratory 2.2.4</u> : State of the art confocal microscopy of biological samples (Dr Stamatis Pagakis) <u>Laboratory 2.3.1</u> : Protein separation by two-dimensional electrophoresis (Dr Antonia Vlahou) <u>Laboratory 2.3.2</u> : Mass spectrometry (Dr Spiros D. Garbis) <u>Laboratory 2.3.3</u> : Fabrication of protein microarrays using nanoplotter (Dr George Tsangaris) <u>Laboratory 2.3.6</u> : Bioinformatics laboratory (Dr Sophia Kossida)
<b>17:15-17:30</b>	<b>Coffee Break</b>
<b>17:30-19:30</b> <b>(shift 4)</b>	<u>Laboratory 2.2.4</u> : State of the art confocal microscopy of biological samples (Dr Stamatis Pagakis) <u>Laboratory 2.3.1</u> : Protein separation by two-dimensional electrophoresis (Dr Antonia Vlahou) <u>Laboratory 2.3.2</u> : Mass spectrometry (Dr Spiros D. Garbis) <u>Laboratory 2.3.3</u> : Fabrication of protein microarrays using nanoplotter (Dr George Tsangaris) <u>Laboratory 2.3.6</u> : Bioinformatics laboratory (Dr Sophia Kossida)

## THURSDAY , 3 JULY – NCSR “DEMOKRITOS”

<b>09:00-10:30</b>	<u>Lecture 2.1.1</u> : Conventional patterning schemes for hard substrates for bioanalytical microdevices (Dr Evangelos Gogolides)
<b>10:30-11:15</b>	<u>Lecture 2.1.2</u> : Microfabrication technologies for plastic analytical microfluidics Part 1 - (Dr Angeliki Tserepi)
<b>11:15-11:30</b>	<b>Coffee Break</b>
<b>11:30-12:15</b>	<u>Lecture 2.1.2</u> : Microfabrication technologies for plastic analytical microfluidics Part 2 - (Dr Angeliki Tserepi)
<b>12:15-13:45</b>	<u>Lecture 2.1.3</u> : Patterning of biomolecules and other biological substances (Dr Panagiotis Argitis)
<b>13:45-14:45</b>	<b>Lunch break</b>

14:45-16:15	Lecture 2.1.4: Molecular bioelectronics (Dr Eleni Makarona)
16:15-16:30	<i>Coffee Break</i>
16:30-18:00	Lecture 3.1: Principles of Integrated Biosensing Devices (Dr Konst. Misiakos)

### FRIDAY , 4 JULY – NCSR “DEMOKRITOS”

09:00-10:30	Lecture 3.3: Acoustic wave sensors: from device fabrication to biological applications ( <b>Invited:</b> Dr Elektra Gizeli)
10:30-10:45	<i>Coffee Break</i>
10:45-14:15 (shift 1)	<p><u>Laboratory 2.1.1:</u> Fabrication of microfluidic devices on plastic substrates by Soft lithography (Dr Angeliki Tserepi)</p> <p><u>Laboratory 2.1.2:</u> Fabrication of plastic microfluidic devices by Lithography and deep polymer plasma etching techniques (Dr Evangelos Gogolides)</p> <p><u>Laboratory 2.1.3:</u> Electrical characterization of tunnelling devices based on organic molecules or biomolecules (Dr Eleni Makarona, Dr Dimitris Velessiotis)</p> <p><u>Laboratory 2.3.4+2.3.5:</u> Fabrication of protein microarrays using lithography (Dr Antonis Douvas) Fluorescence detection of protein arrays (Dr Panagiota Petrou)</p> <p><u>Laboratory 3.1:</u> Operation of a lab-on-a-chip optical device using model assays and real time measurements (Dr Konstantinos Misiakos)</p>

### MONDAY , 7 JULY – NCSR “DEMOKRITOS”

09:00-11:30 (shift 2)	<p><u>Laboratory 2.1.1:</u> Fabrication of microfluidic devices on plastic substrates by lithographic techniques (Dr Angeliki Tserepi)</p> <p><u>Laboratory 2.1.2:</u> Fabrication of plastic microfluidic devices by Lithography and deep polymer plasma etching techniques (Dr Evangelos Gogolides)</p> <p><u>Laboratory 2.1.3:</u> Electrical characterization of tunnelling devices based on organic molecules or biomolecules (Dr Eleni Makarona, Dr Dimitris Velessiotis)</p> <p><u>Laboratory 2.3.4+2.3.5:</u> Fabrication of protein microarrays using lithography (Dr Antonis Douvas) Fluorescence detection of protein arrays (Dr Panagiota Petrou)</p> <p><u>Laboratory 3.1:</u> Operation of a lab-on-a-chip optical device using model assays and real time measurements (Dr Konstantinos Misiakos)</p>
11:30-12:00	<i>Coffee break</i>
12:00-14:30 (shift 3)	<p><u>Laboratory 2.1.1:</u> Fabrication of microfluidic devices on plastic substrates by lithographic techniques (Dr Angeliki Tserepi)</p> <p><u>Laboratory 2.1.2:</u> Fabrication of plastic microfluidic devices by Lithography and deep polymer plasma etching techniques (Dr Evangelos Gogolides)</p> <p><u>Laboratory 2.1.3:</u> Electrical characterization of tunnelling devices based on organic molecules or biomolecules (Dr Eleni Makarona, Dr Dimitris Velessiotis)</p> <p><u>Laboratory 2.3.4+2.3.5:</u> Fabrication of protein microarrays using lithography (Dr Antonis Douvas) Fluorescence detection of protein arrays (Dr Panagiota Petrou)</p> <p><u>Laboratory 3.1:</u> Operation of a lab-on-a-chip optical device using model assays and real time measurements (Dr Konstantinos Misiakos)</p>

<b>14:30-15:30</b>	<b>Lunch break</b>
<b>15:30-18:00</b> <b>(shift 4)</b>	<p><u>Laboratory 2.1.1:</u> Fabrication of microfluidic devices on plastic substrates by lithographic techniques (Dr Angeliki Tserepi)</p> <p><u>Laboratory 2.1.2:</u> Fabrication of plastic microfluidic devices by Lithography and deep polymer plasma etching techniques (Dr Evangelos Gogolides)</p> <p><u>Laboratory 2.1.3:</u> Electrical characterization of tunnelling devices based on organic molecules or biomolecules (Dr Eleni Makarona, Dr Dimitris Velessiotis)</p> <p><u>Laboratory 2.3.4+2.3.5:</u> Fabrication of protein microarrays using lithography (Dr Antonis Douvas) Fluorescence detection of protein arrays (Dr Panagiota Petrou)</p> <p><u>Laboratory 3.1:</u> Operation of a lab-on-a-chip optical device using model assays and real time measurements (Dr Konstantinos Misiakos)</p>

## TUESDAY , 8 JULY – NCSR “DEMOKRITOS”

<b>09:00-09:45</b>	<u>Lecture 2.2.1:</u> Targeting RNA with small molecules: a Pharmaceutical Industry Study (Dr Dionysios Vourloumis)
<b>09:45-11:00</b>	<u>Lecture 2.2.2 and 2.2.3:</u> Drug Delivery and Targeting Systems - Focus on Liposomes ( <b>Invited:</b> Prof. Sophia Antimisiaris)
<b>11:00-11:15</b>	<b>Coffee Break</b>
<b>11:15-12:00</b>	<u>Lecture 2.2.5:</u> Magnetic nanoparticles for bioapplications (Dr Ioannis Rabias)
<b>12:00-12:45</b>	<u>Lecture 2.2.6:</u> Principles of Biomimetics in Core Nanotechnologies (Prof. Georgia C. Papaefthymiou)
<b>12:45-13:45</b>	<b>Lunch break</b>
<b>13:45-14:30</b>	<u>Lecture 2.2.4:</u> Controlled positioning of biomolecules using lasers ( <b>Invited:</b> Dr Maria Farsari)
<b>14:30-15:15</b>	<u>Lecture 2.3.5:</u> Metabolomics in the Post-Genomic Era ( <b>Invited:</b> Dr Maria I. Klapa)
<b>15:15-16:30</b>	<u>Lecture 2.2.2 and 2.2.3:</u> Drug Release and Delivery Systems - Methods (Dr Irene Mavridis, Dr Konstantina Yannakopoulou)
<b>16:30</b> <b>20:00</b>	<i>Excursion to Cape Sounion</i> <b>Dinner</b>

## WEDNESDAY , 9 JULY – NCSR “DEMOKRITOS”

<b>10:15-13:45</b> <b>(shift 1)</b>	<p><u>Laboratory 2.2.1:</u> Drug inclusion in cyclodextrins: monitoring in situ by NMR spectroscopy, X-ray diffraction characterisation of drug inclusion and 3-D visualisation (Dr K. Yannakopoulou, Dr E. Saridakis, Dr A. Paulidou)</p> <p><u>Laboratory 2.2.2+2.2.3:</u> • Liposomes: preparation and characterisation by dynamic light scattering and <math>\zeta</math>-potential • Video enhanced optical microscopy and Atomic Force Microscopy of Liposomes (Dr Dimitrios Tsiourvas, Dr Zili Sideratou)</p> <p><u>Laboratory 2.2.5+2.2.6:</u> Magnetic nanomaterials for bioapplications Magnetic hyperthermia for Biomedical applications (Dr Ioannis Rabias) Determining Magnetic Anisotropy at the Nanoscale Case study: Horse Spleen Ferritin STEM, Mössbauer and SQUID measurements (Prof. Georgia C. Papaefthymiou)</p> <p><u>Laboratory 3.2:</u> Demonstration of a capillary fluoroimmunosensor (Dr Sotirios Kakabakos)</p>
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<b>13:45-14:45</b>	<b>Lunch break</b>
<b>14:45-18:00</b> <b>(shift 2)</b>	<p><u>Laboratory 2.2.1:</u> Drug inclusion in cyclodextrins: monitoring in situ by NMR spectroscopy, X-ray diffraction characterisation of drug inclusion and 3-D visualisation (Dr K. Yannakopoulou, Dr E. Saridakis, Dr A. Paulidou)</p> <p><u>Laboratory 2.2.2+2.2.3:</u> • Liposomes: preparation and characterisation by dynamic light scattering and <math>\zeta</math>-potential • Video enhanced optical microscopy and Atomic Force Microscopy of Liposomes (Dr Dimitrios Tsiourvas)</p> <p><u>Laboratory 2.2.5+2.2.6:</u> Magnetic nanomaterials for bioapplications Magnetic hyperthermia for Biomedical applications (Dr Ioannis Rabias) Determining Magnetic Anisotropy at the Nanoscale Case study: Horse Spleen Ferritin STEM, Mössbauer and SQUID measurements (Prof. Georgia C. Papaefthymiou)</p> <p><u>Laboratory 3.2:</u> Demonstration of a capillary fluoroimmunosensor (Dr Sotirios Kakabakos)</p>

<b>THURSDAY , 10 JULY – NCSR “DEMOKRITOS”</b>
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<b>09:00-12:30</b> <b>(shift 3)</b>	<p><u>Laboratory 2.2.1:</u> Drug inclusion in cyclodextrins: monitoring in situ by NMR spectroscopy, X-ray diffraction characterisation of drug inclusion and 3-D visualisation (Dr K. Yannakopoulou, Dr E. Saridakis, Dr A. Paulidou)</p> <p><u>Laboratory 2.2.2+2.2.3:</u> • Liposomes: preparation and characterisation by dynamic light scattering and <math>\zeta</math>-potential • Video enhanced optical microscopy and Atomic Force Microscopy of Liposomes (Dr Dimitrios Tsiourvas)</p> <p><u>Laboratory 2.2.5+2.2.6:</u> Magnetic nanomaterials for bioapplications Magnetic hyperthermia for Biomedical applications (Dr Ioannis Rabias) Determining Magnetic Anisotropy at the Nanoscale Case study: Horse Spleen Ferritin STEM, Mössbauer and SQUID measurements (Prof. Georgia C. Papaefthymiou)</p> <p><u>Laboratory 3.2:</u> Demonstration of a capillary fluoroimmunosensor (Dr Sotirios Kakabakos)</p>
<b>12:30-13:00</b>	<b>Closing ceremony</b>
<b>13:00-14:00</b>	<b>Lunch</b>